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FILE COVERS 1974 TO 13 Jul 2001 (20010713/ED)

=> E SASAKI I, 1979/RE
E1 8 SASAKI I, 1976, V33, P162,
KOBUNSHI RONBUNSHU/RE
E2 1 SASAKI I, 1976, V41, P181,
SCI PEST CONTR/RE
E3 0 --> SASAKI I, 1979/RE
E4 1 SASAKI I, 1979, V86, P1537,
BIOCHEM/RE
E5 11 SASAKI I, 1979, V86, P1537,
J BIOCH/RE
E6 1 SASAKI I, 1979, V86, P1537,
J BIOCHEM TOKYO/RE
E7 1 SASAKI I, 1980, 6TH ECOD/RE
E8 1 SASAKI I, 1980, 6TH EUR C
OPT COMM Y/RE
E9 1 SASAKI I, 1980, EUROPEAN C
OPTICAL F/RE
E10 2 SASAKI I, 1980, P140, 6TH
EUR C OPT COMM Y/RE
E11 8 SASAKI I, 1980, P140, 6TH P
EUR C OPT COMM/RE
E12 38 SASAKI I, 1980, V16, P219,
ELECTRON LETT/RE

=> S E4-6
1 "SASAKI I, 1979, V86, P1537,
BIOCHEM"/RE
("SASAKI I, 1979, V86, P1537,
BIOCHEM"/RE)
11 "SASAKI I, 1979, V86, P1537, J
BIOCH"/RE
("SASAKI I, 1979, V86, P1537,
J BIOCH"/RE)
1 "SASAKI I, 1979, V86, P1537, J
BIOCHEM TOKYO"/RE
("SASAKI I, 1979, V86, P1537,
J BIOCHEM TOKYO"/RE)
L1 13 ("SASAKI I, 1979, V86, P1537,
BIOCHEM"/RE OR "SASAKI I, 1979,
V86, P1537, J BIOCH"/RE OR
"SASAKI I, 1979, V86, P1537, J BIOCHE
M TOKYO"/RE)

=> E SASAKI I, 1982/RE
E1 1 SASAKI I, 1981, V38, P75,
KONBUNSHI RONBUNSHU/RE
E2 1 SASAKI I, 1981, V7, P90,
AQUICULTURE/RE

E3 0 --> SASAKI I, 1982/RE
E4 1 SASAKI I, 1982, APR OFC 82
PHOEN/RE
E5 1 SASAKI I, 1982, P30, P OFC
PHOENIX/RE
E6 1 SASAKI I, 1982, P341, NOUV
J CHIM/RE
E7 1 SASAKI I, 1982, THESIS U
SOUTHAMPTON/RE
E8 1 SASAKI I, 1982, V21, APPL
OPTICS/RE
E9 1 SASAKI I, 1982, V21, P4246,
APPL OPTICS/RE
E10 1 SASAKI I, 1982, V21, P4256,
APPL OPTICS/RE
E11 1 SASAKI I, 1982, V24, P495,
EXP BRAIN RES/RE
E12 7 SASAKI I, 1982, V6, P341,
NOUV J CHIM/RE

=> E
E13 6 SASAKI I, 1982, V91, P1555,
J BIOCH/RE
E14 3 SASAKI I, 1982, V91, P1555,
J BIOCHEM-TOKYO/RE
E15 15 SASAKI I, 1982, V91, P211,
J BIOCH/RE
E16 1 SASAKI I, 1983, THESIS U
PARIS SUD O/RE
E17 3 SASAKI I, 1984, V4, P237,
NOUV J CHIM/RE
E18 1 SASAKI I, 1984, V8, P237,
NOUV J CHIM/RE
E19 1 SASAKI I, 1984, V9, P385, J
MAGN SOC JAPAN/RE
E20 1 SASAKI I, 1985, P ISCAS
85/RE
E21 2 SASAKI I, 1985, P1633, P
ISCAS 85/RE
E22 9 SASAKI I, 1985, V332, P237,
J CHROMATOGR/RE
E23 1 SASAKI I, 1985, V68, P842,
T IECE C/RE
E24 1 SASAKI I, 1985, V68, P842,
T IECE J/RE

=> S E13-14
6 "SASAKI I, 1982, V91, P1555, J
BIOCH"/RE
("SASAKI I, 1982, V91, P1555,
J BIOCH"/RE)
3 "SASAKI I, 1982, V91, P1555, J
BIOCHEM-TOKYO"/RE
("SASAKI I, 1982, V91, P1555,
J BIOCHEM-TOKYO"/RE)
L2 9 ("SASAKI I, 1982, V91, P1555, J
BIOCH"/RE OR "SASAKI I, 1982,
V91, P1555, J BIOCHEM-
TOKYO"/RE)

=> S L1 OR L2
L3 20 L1 OR L2

=> D BIB AB 1-20

L3 ANSWER 1 OF 20 SCISEARCH COPYRIGHT 2001
ISI (R)
AN 2001:220451 SCISEARCH
GA The Genuine Article (R) Number: 407JQ
TI Purification and partial characterization
of a cholesterol oxidase from

Streptomyces fradiae
 AU Yazdi M T (Reprint); Zahraei M; Aghaepour
 K; Kamranpour N
 CS Tehran Univ Med Sci, Coll Pharm, Dept
 Biotechnol, Tehran, Iran (Reprint);
 Tehran Univ Med Sci, Coll Med, Dept
 Biochem, Tehran, Iran
 CYA Iran
 SO ENZYME AND MICROBIAL TECHNOLOGY, (8 MAR
 2001) Vol. 28, No. 4-5, pp.
 410-414.
 Publisher: ELSEVIER SCIENCE INC, 655
 AVENUE OF THE AMERICAS, NEW YORK, NY
 10010 USA.
 ISSN: 0141-0229.
 DT Article; Journal
 LA English
 REC Reference Count: 28
 *ABSTRACT IS AVAILABLE IN THE ALL AND
 IALL FORMATS*
 AB An extracellular cholesterol oxidase
 from Streptomyces fradiae (PTCC
 1121) was purified in one step using
 DEAE-Sephadex. The purified enzyme
 had a molecular weight of 60 kDa. The
 optimum pH and temperature for
 activity was found to be 7 and 70
 degreesC, respectively. This cholesterol
 oxidase was stable in pHs between 4-10 at
 4 degreesC until 4 h. Thermal
 stability experiments showed that it has
 high stability and retains its
 full activity at 50 degreesC for 90 min.
 K-m value for cholesterol oxidase
 was obtained to be about 7.06×10^{-5}
 Mol. (C) 2001 Elsevier Science Inc.
 All rights reserved.

L3 ANSWER 2 OF 20 SCISEARCH COPYRIGHT 2001
 ISI (R)
 AN 2000:287062 SCISEARCH
 GA The Genuine Article (R) Number: 302HA
 TI Salivary amylase activity of the
 phlebotomine sand fly, Lutzomyia
 longipalpis
 AU Ribeiro J M C (Reprint); Rowton E D;
 Charlamb R
 CS NIAID, SECT MED ENTOMOL, PARASIT DIS LAB,
 NIH, BLDG 4, ROOM 126, 4 CTR DR,
 MSC-0425, BETHESDA, MD 20892 (Reprint);
 WALTER REED ARMY MED CTR, WALTER
 REED ARMY INST RES, DEPT ENTOMOL,
 WASHINGTON, DC 20307
 CYA USA
 SO INSECT BIOCHEMISTRY AND MOLECULAR
 BIOLOGY, (APR 2000) Vol. 30, No. 4, pp.
 271-277.
 Publisher: PERGAMON-ELSEVIER SCIENCE LTD,
 THE BOULEVARD, LANGFORD LANE,
 KIDLINGTON, OXFORD OX5 1GB, ENGLAND.
 ISSN: 0965-1748.
 DT Article; Journal
 FS LIFE; AGRI
 LA English
 REC Reference Count: 28
 *ABSTRACT IS AVAILABLE IN THE ALL AND
 IALL FORMATS*
 AB Both male and female adult stages of
 the sand fly Lutzomyia longipalpis
 have detectable amylase activity in their
 salivary glands, as indicated by

formation of p-nitrophenyl-alpha-D-
 maltoside from p-nitrophenyl-alpha-D-
 octoside and by hydrolysis of 4-
 nitrophenyl-alpha-D-maltoheptaoside-4,
 6,-O-ethylidene. No salivary alpha-
 glucosidase was detected. Amylase
 activity was also found in the crop and
 midgut of female flies, although
 in a smaller amount. Salivary amylase is
 significantly reduced from the
 salivary glands immediately after a blood
 meal, as is the case with
 salivary alpha-glucosidases in
 mosquitoes. Presence of salivary gland
 amylase in these sand flies, and absence
 of salivary alpha-glucosidase,
 indicates that in nature these insects
 may have a significant intake of
 carbohydrates in the form of starch, as
 suggested by their plant-feeding
 behavior, previously demonstrated by
 Schlein and Warburg (Schlein, Y.,
 Warburg, A., 1986. Phytophagy and the
 feeding cycle of Phlebotomus
 papatasi (Diptera: Psychodidae) under
 experimental conditions. Journal of
 Medical Entomology 23, 11-15), and
 Alexander and Usma (Alexander, B.,
 Usma, M.C., 1994. Potential sources of
 sugar for the phlebotomine sandfly
 Lutzomyia youngi (Diptera: Psychodidae)
 in a Columbia coffee plantation.
 Ann. Trop. Med. Parasitol. 88, 543-549).
 Published by Elsevier Science
 Ltd.

L3 ANSWER 3 OF 20 SCISEARCH COPYRIGHT 2001
 ISI (R)
 AN 1999:920078 SCISEARCH
 GA The Genuine Article (R) Number: 257WZ
 TI Capture of human Fab fragments by
 expanded bed adsorption with a mixed
 mode adsorbent
 AU Hansen M B (Reprint); Lihme A; Spitali M;
 King D
 CS UPFRONT CHROMATOGRAPHY, DK-2100 COPENHAGEN,
 DENMARK; CELLTECH THERAPEUT,
 SLOUGH, BERKS, ENGLAND
 CYA DENMARK; ENGLAND
 SO BIOSEPARATION, (SEP 1998) Vol. 8, No. 1-
 5, pp. 189-193.
 Publisher: KLUWER ACADEMIC PUBL,
 SPUIBOULEVARD 50, PO BOX 17, 3300 AA
 DORDRECHT, NETHERLANDS.
 ISSN: 0923-179X.
 DT Article; Journal
 LA English
 REC Reference Count: 22
 *ABSTRACT IS AVAILABLE IN THE ALL AND
 IALL FORMATS*
 AB A novel group of mixed mode adsorbents
 has been developed for
 purification of monoclonal and polyclonal
 antibodies from a broad range of
 raw materials such as hybridoma cell
 culture, ascites fluid, animal sera,
 milk, whey and egg yolk. The aim of this
 study was to determine whether
 such mixed mode adsorbents were also
 useful for the recovery of

recombinant proteins from microbial feedstocks. This paper describes the performance of one of these adsorbents for expanded bed capture of a human Fab fragment from recombinant *E. Coli* cell extracts.

It is concluded that the mixed mode adsorbent binds the Fab fragment efficiently from crude extracts without any requirement for preconditioning the extract by for example de-salting or dilution. The capacity of the mixed mode adsorbent is approx. 12 mg Fab/ml matrix.

The novel mixed mode adsorbent can be useful during production of highly purified Fab fragments as the first step in a purification scheme. In this respect the mixed mode adsorbent is advantageous over alternative commercially available ion-exchange materials which require pre-conditioning of cell extract for Fab capture. Together with the concentration and clarification effect a significant enrichment of the Fab fragment is obtained in one single high yield operation.

L3 ANSWER 4 OF 20 SCISEARCH COPYRIGHT 2001
ISI (R)
AN 1998:894226 SCISEARCH
GA The Genuine Article (R) Number: 139XY
TI Characterization of cytochrome c-556 from the purple phototrophic bacterium *Rhodobacter capsulatus* as a cytochrome-c peroxidase
AU Hu W; DeSmet L; VanDriessche G; Bartsch R G; Meyer T E; Cusanovich M A; VanBeeumen J (Reprint)
CS STATE UNIV GHENT, LAB EIWITBIOCHEM
EIWITENG, LEDEGANCKSTR 35, B-9000 GHENT, BELGIUM (Reprint); STATE UNIV GHENT, DEPT BIOCHEM PHYSIOL & MICROBIOL, LAB PROT BIOCHEM & PROT ENGN, GHENT, BELGIUM; UNIV ARIZONA, DEPT BIOCHEM, TUCSON, AZ 85721
CYA BELGIUM; USA
SO EUROPEAN JOURNAL OF BIOCHEMISTRY, (15 NOV 1998) Vol. 258, No. 1, pp. 29-36.
Publisher: SPRINGER VERLAG, 175 FIFTH AVE, NEW YORK, NY 10010.
ISSN: 0014-2956.
DT Article; Journal
FS LIFE
LA English
REC Reference Count: 36
ABSTRACT IS AVAILABLE IN THE ALL AND IALL FORMATS

AB A cytochrome c-556 was purified from *Rhodobacter capsulatus* and the complete amino acid sequence was determined. It contains 328 amino acid residues and two typical heme-binding sites at cysteine residues 54 and 57 and at residues 200 and 203. It is homologous to the family of bacterial cytochrome c peroxidases (BCCP) with 69% identity to *Paracoccus*

denitrificans BCCP and 60% identity to *Pseudomonas aeruginosa* BCCP for which there is a three-dimensional structure. There is lesser similarity to the *mauG* gene products from methylotrophic bacteria which are thought to be involved in biosynthesis of the quinone cofactor of methylamine dehydrogenase. Translated genes from *Escherichia coli* and *Helicobacter pylori* are also related to the bacterial cytochrome c peroxidases. The divergence of this family of proteins is reflected in the fact that the reported sixth heme ligands are not conserved, except in *Pseudomonas*, *Rhodobacter* and *Paracoccus*. This suggests that homologs of BCCP may fold differently and/or may not have the same enzymatic activity as the prototypic protein from *Ps. aeruginosa*. We found that the *Rb. capsulatus* BCCP is active with both *Rb. capsulatus* cytochrome c, and with horse cytochrome c as substrates (K-m values 60 μ M and 6 μ M, respectively). The turnover number was 40 s⁻¹ and the K-m for peroxide was 33 μ M. We have thus confirmed that the *Rb. capsulatus* protein is a cytochrome c peroxidase.

L3 ANSWER 5 OF 20 SCISEARCH COPYRIGHT 2001
ISI (R)
AN 1998:609664 SCISEARCH
GA The Genuine Article (R) Number: 106XH
TI Hydrophobic charge induction chromatography: salt independent protein adsorption and facile elution With aqueous buffers
AU Burton S C; Harding D R K (Reprint)
CS MASSEY UNIV, DEPT CHEM, PALMERSTON NORTH, NEW ZEALAND (Reprint); MASSEY UNIV, DEPT CHEM, PALMERSTON NORTH, NEW ZEALAND
CYA NEW ZEALAND
SO JOURNAL OF CHROMATOGRAPHY A, (24 JUL 1998) Vol. 814, No. 1-2, pp. 71-81.
Publisher: ELSEVIER SCIENCE BV, PO BOX 211, 1000 AE AMSTERDAM, NETHERLANDS.
ISSN: 0021-9673.
DT Article; Journal
FS PHYS; LIFE
LA English
REC Reference Count: 27
ABSTRACT IS AVAILABLE IN THE ALL AND IALL FORMATS

AB A new form of protein chromatography, hydrophobic charge induction, is described. Matrices prepared by attachment of weak acid and base ligands were uncharged at adsorption pH. At low ligand densities, protein adsorption was typically promoted with lyotropic salts. At higher ligand densities, chymosin, chymotrypsinogen and lysozyme were adsorbed independently of ionic strength. A pH change released the electrostatic

QD241,
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potential of the matrix and weakened hydrophobic interactions, inducing elution. Matrix hydrophobicity and titration range could be matched to protein requirements by ligand choice and density. Both adsorption and elution could be carried out within the pH 5-9 range. (C) 1998 Elsevier Science B.V. All rights reserved.

L3 ANSWER 6 OF 20 SCISEARCH COPYRIGHT 2001
ISI (R)
AN 97:681234 SCISEARCH
GA The Genuine Article (R) Number: XU956
TI One step purification of chymosin by mixed mode chromatography
AU Burton S C; Haggarty N W; Harding D R R (Reprint)
CS MASSEY UNIV, DEPT CHEM, PRIVATE BAG 11222, PALMERSTON NORTH, NEW ZEALAND (Reprint); MASSEY UNIV, DEPT CHEM, PALMERSTON NORTH, NEW ZEALAND
CYA NEW ZEALAND
SO BIOTECHNOLOGY AND BIOENGINEERING, (5 OCT 1997) Vol. 56, No. 1, pp. 45-55.
Publisher: JOHN WILEY & SONS INC, 605 THIRD AVE, NEW YORK, NY 10158-0012.
ISSN: 0006-3592.
DT Article; Journal
FS LIFE; AGRI
LA English
REC Reference Count: 37
ABSTRACT IS AVAILABLE IN THE ALL AND IALL FORMATS

AB Mixed mode Sepharose and Perloza bead cellulose matrices were prepared using various chemistries. These matrices contained hydrophobic (aliphatic and/or aromatic) and ionic (carboxylate or alkylamine) groups. Hydrophobic amine ligands were attached to epichlorohydrin activated Sepharose (mixed mode amine matrices). Hexylamine, aminophenylpropanediol and phenylethylamine were the preferred ligands, on the basis of cost and performance. Other mixed mode matrices were produced by incomplete attachment (0-80%) of the same amine ligands to carboxylate matrices. The best results were obtained using unmodified or partially ligand-modified aminocaproic acid Sepharose and Perloza. High ligand densities were used, resulting in high capacity. Furthermore, chymosin was adsorbed at high and low ionic strengths, which reduced sample preparation requirements.

Chymosin, essentially homogeneous by electrophoresis, was recovered by a small pH change. The methods described were simple, efficient, inexpensive and provided very good resolution of chymosin from a crude recombinant source. The carboxylate matrices had the best combination of capacity and regeneration properties. The performance of Sepharose and Perloza carboxylate matrices was similar, but higher capacities were found for the

latter. Because it is cheaper and can be used at higher flow rates, Perloza should be better suited to large scale application. High capacity chymosin adsorption was found with carboxymethyl ion exchange matrices, but low ionic strength was essential for adsorption and the purity was inferior to that of the mixed mode matrices. (C) 1997 John Wiley & Sons, Inc.

L3 ANSWER 7 OF 20 SCISEARCH COPYRIGHT 2001
ISI (R)
AN 94:358130 SCISEARCH
GA The Genuine Article (R) Number: NP630
TI A NEW MICROORGANISM PRODUCING A GLUCOSYL TRANSFER ENZYME TO POLYPHENOLS
AU FUNAYAMA M (Reprint); ARAKAWA H; YAMAMOTO R; NISHINO T; SHIN T; MURAO S
CS KURABO IND LTD, TECH RES LAB, 14-5 SHIMOKIDA CHO, NEYAGAWA, OSAKA 572, JAPAN (Reprint); KUMAMOTO INST TECHNOL, FAC ENGN, DEPT APPL MICROBIAL TECHNOL, KUMAMOTO 860, JAPAN
CYA JAPAN
SO BIOSCIENCE BIOTECHNOLOGY AND BIOCHEMISTRY, (MAY 1994) Vol. 58, No. 5, pp. 817-821.
ISSN: 0916-8451.
DT Article; Journal
FS LIFE; AGRI
LA ENGLISH
REC Reference Count: 14
ABSTRACT IS AVAILABLE IN THE ALL AND IALL FORMATS

AB A microorganism producing a glucosyl transfer enzyme to hydroquinone was isolated from soil and identified as *Bacillus subtilis* according to its taxonomical characteristics. The enzyme (GSase) was purified from the culture filtrate by column chromatographies, including affinity chromatography using Amylostatin-immobilized Sepharose 4B. The final preparation showed a single band on SDS polyacrylamide gel electrophoresis, the molecular weight being 67 kDa. Its optimum pH for starch dextrinization was 7, while that for glucosyl transferring activity was 6, pH stability was 5-8, and isoelectric point was 5.1. GSase was not activated by Ca^{2+} . It used malto-oligosaccharides and dextrin as well as soluble starch more effectively than maltose as glucose donors. It did not catalyze cyclodextrination from starch. GSase glucosylated various polyphenols, such as dihydroxy benzenes, hydroxy benzyl alcohols, phloroglucinol, (+)catechin, kojic acid, dihydroxy benzoic acids, caffeic acid, and gallic acid.

L3 ANSWER 8 OF 20 SCISEARCH COPYRIGHT 2001
ISI (R)
AN 92:35852 SCISEARCH
GA The Genuine Article (R) Number: GY133

TI ENZYME-CATALYZED OXIDATION OF CHOLESTEROL
IN PHYSICALLY CHARACTERIZED
WATER-IN-OIL MICROEMULSIONS

AU HEDSTROM G; SLOTT J P (Reprint);
MOLANDER O; ROSENHOLM J B
CS ABO AKAD UNIV, DEPT BIOCHEM & PHARM, SF-
20500 TURKU, FINLAND; ABO AKAD
UNIV, DEPT PHYS CHEM, SF-20500 TURKU,
FINLAND

CYA FINLAND
SO BIOTECHNOLOGY AND BIOENGINEERING, (20 JAN
1992) Vol. 39, No. 2, pp.

218-224.
ISSN: 0006-3597.

DT Article; Journal

FS LIFE; AGRI

LA ENGLISH

REC Reference Count: 24

*ABSTRACT IS AVAILABLE IN THE ALL AND
IAL FORMATS*

AB The enzymatic conversion of
cholesterol to cholestenone by cholesterol
oxidase (Brevibacterium sp.) in reversed
micelles in a system composed of
AOT/isooctane/water/cholesterol has been
examined. The catalytic activity
of the enzyme was correlated with the
physicochemical properties of water
in water-in-oil (w/o) microemulsion
systems. In a system consisting of 3
wt % AOT in isooctane, reversed micelles
started to form as the
[H₂O]/[AOT] (e.g., the w/o) ratio
increased above 4-5. The formation of
reversed micelles with a core of neat
(bulk) water was verified from
determinations of both the partial molar
volume of water and the scissors
vibration of water [with Fourier
transform infrared (FTIR) spectroscopy]
in the w/o microemulsion systems. A plot
of enzyme activity vs. w/o
indicated that the hydration of enzyme
molecules per se was not sufficient
to give rise to catalytic activity.
Instead, it appeared that the
formation of an aqueous micellar core was
necessary for full activation of
the enzyme. Based on micelle size
distribution analysis, it was estimated
that about one micelle per one thousand
contained an enzyme molecule.
Since the apparent reaction rate could be
markedly enhanced by increasing
the enzyme/water ratio, we conclude that
the number of enzyme-containing
micelles was an important rate-limiting
factor in the system.

L3 ANSWER 9 OF 20 SCISEARCH COPYRIGHT 2001
ISI (R)

AN 88:147205 SCISEARCH

GA The Genuine Article (R) Number: M4528

TI CHOLESTEROL CONVERSION TO DELTA-4-
CHOLESTENONE BY CHOLESTEROL OXIDASE IN
POLYPHASIC SYSTEMS - EXTENSION TO THE
SELECTIVE OXIDATION OF
7-BETA-HYDROXYCHOLESTEROL

AU LEE K M (Reprint); BIELLMANN J F

CS UNIV STRASBOURG 1, INST CHIM, CHIM ORGAN
BIOL LAB, UNITE 31, 1 RUE BLAISE

PASCAL, F-67008 STRASBOURG, FRANCE

(Reprint)

CYA FRANCE

SO TETRAHEDRON, (1988) Vol. 44, No. 4, pp.
1135-1139.

DT Article; Journal

FS PHYS; LIFE

LA ENGLISH

REC Reference Count: 31

L3 ANSWER 10 OF 20 SCISEARCH COPYRIGHT

2001 ISI (R)

AN 87:523362 SCISEARCH

GA The Genuine Article (R) Number: J9748

TI CRYSTALLIZATION AND MOLECULAR-PROPERTIES
OF D-2-HYDROXYISOCAPROATE

DEHYDROGENASE FROM LACTOBACILLUS-CASEI

AU KALLWASS H; TSAI H (Reprint); SCHUTTE H

CS GESELL BIOTECHNOL FORSCH MBH,
ENZYMTECHNOL ABT, MASCHERODER WEG 1, D-3300
BRUNSWICK, FED REP GER

CYA GERMANY

SO FEMS MICROBIOLOGY LETTERS, (1987) Vol.
43, No. 3, pp. 263-267.

DT Article; Journal

FS LIFE

LA ENGLISH

REC Reference Count: 20

L3 ANSWER 11 OF 20 SCISEARCH COPYRIGHT

2001 ISI (R)

AN 86:540968 SCISEARCH

GA The Genuine Article (R) Number: E1197

TI CHOLESTEROL OXIDASE IN MICROEMULSION -
ENZYMATIC-ACTIVITY ON A SUBSTRATE

OF LOW WATER SOLUBILITY AND INACTIVATION
BY HYDROGEN-PEROXIDE

AU LEE K M (Reprint); BIELLMANN J F

CS UNIV STRASBOURG 1, INST CHIM, CNRS, CHIM
ORGAN BIOL LAB, F-67008

STRASBOURG, FRANCE (Reprint)

CYA FRANCE

SO BIOORGANIC CHEMISTRY, (1986) Vol. 14, No.
3, pp. 262-273.

DT Article; Journal

FS PHYS; LIFE

LA ENGLISH

REC Reference Count: 25

L3 ANSWER 12 OF 20 SCISEARCH COPYRIGHT

2001 ISI (R)

AN 86:276749 SCISEARCH

GA The Genuine Article (R) Number: C2081

TI POLYSACCHARIDE LYASES

AU LINHARDT R J (Reprint); GALLIHER P M;

COONEY C L

CS UNIV IOWA, COLL PHARM, DIV MED CHEM, IOWA
CITY, IA, 52242 (Reprint);

BIOGEN CORP, CAMBRIDGE, MA, 02139; MIT,
DEPT CHEM ENGN, CAMBRIDGE, MA,
02139

CYA USA

SO APPLIED BIOCHEMISTRY AND BIOTECHNOLOGY,
(1986) Vol. 12, No. 2, pp.
135-176.

DT General Review; Bibliography; Journal

FS LIFE; AGRI

LA ENGLISH

REC Reference Count: 196

2003 AS2
TA

MF

Q9501.858

QD241.74

L3 ANSWER 13 OF 20 SCISEARCH COPYRIGHT
2001 ISI (R) *S583, A37*
AN 83:611739 SCISEARCH
GA The Genuine Article (R) Number: RS814
TI AN ISOMALTOTRIOSE-PRODUCING DEXTRANASE
FROM FLAVOBACTERIUM-SP M-73 - *missing*
PURIFICATION AND PROPERTIES
AU KOBAYASHI M (Reprint); TAKAGI S; SHIOTA
M; MITSUISHI Y; MATSUDA K
CS TOHOKU UNIV, FAC AGR, DEPT AGR CHEM,
SENDAI, MIYAGI 980, JAPAN (Reprint)
CYA JAPAN
SO AGRICULTURAL AND BIOLOGICAL CHEMISTRY,
(1983) Vol. 47, No. 11, pp.
2585-2593.
DT Article; Journal
FS LIFE; AGRI
LA ENGLISH
REC Reference Count: 22

L3 ANSWER 14 OF 20 SCISEARCH COPYRIGHT
2001 ISI (R) *MF*
AN 83:228170 SCISEARCH
GA The Genuine Article (R) Number: QN093
TI COAGULATION OF SKIM MILK WITH PROTEASES
IMMOBILIZED ON HYDROPHOBIC
CARRIERS
AU VOUTSINAS L P (Reprint); NAKAI S
CS UNIV BRITISH COLUMBIA, DEPT FOOD SCI,
VANCOUVER V6T 2A2, BC, CANADA
(Reprint)
CYA CANADA
SO JOURNAL OF DAIRY SCIENCE, (1983) Vol. 66,
No. 4, pp. 694-703.
DT Article; Journal
FS AGRI
LA ENGLISH
REC Reference Count: 37

L3 ANSWER 15 OF 20 SCISEARCH COPYRIGHT ✓
2001 ISI (R)
AN 82:229260 SCISEARCH
GA The Genuine Article (R) Number: NN600
TI HYDROPHOBIC-IONIC CHROMATOGRAPHY - ITS
APPLICATION TO MICROBIAL
GLUCOSE-OXIDASE, HYALURONIDASE,
CHOLESTEROL OXIDASE, AND CHOLESTEROL
ESTERASE
AU SASAKI I (Reprint); GOTOH H; YAMAMOTO R;
TANAKA H; TAKAMI K; YAMASHITA K;
YAMASHITA J; HORIO T
CS OSAKA UNIV, INST PROT RES, DIV ENZYMOL,
SUITA, OSAKA 565, JAPAN (Reprint);
AMANO PHARMACEUT CO LTD, NAGOYA, AICHI
460, JAPAN
CYA JAPAN
SO JOURNAL OF BIOCHEMISTRY, (1982) Vol. 91,
No. 5, pp. 1555-1561.
DT Article; Journal
FS LIFE
LA ENGLISH
REC Reference Count: 8

L3 ANSWER 16 OF 20 SCISEARCH COPYRIGHT
2001 ISI (R) *QP501, J6*
AN 82:49286 SCISEARCH
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